

# Enzyme Synergies in the Hydrolysis of AFEX Pretreated Biomass

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# Scope of Presentation

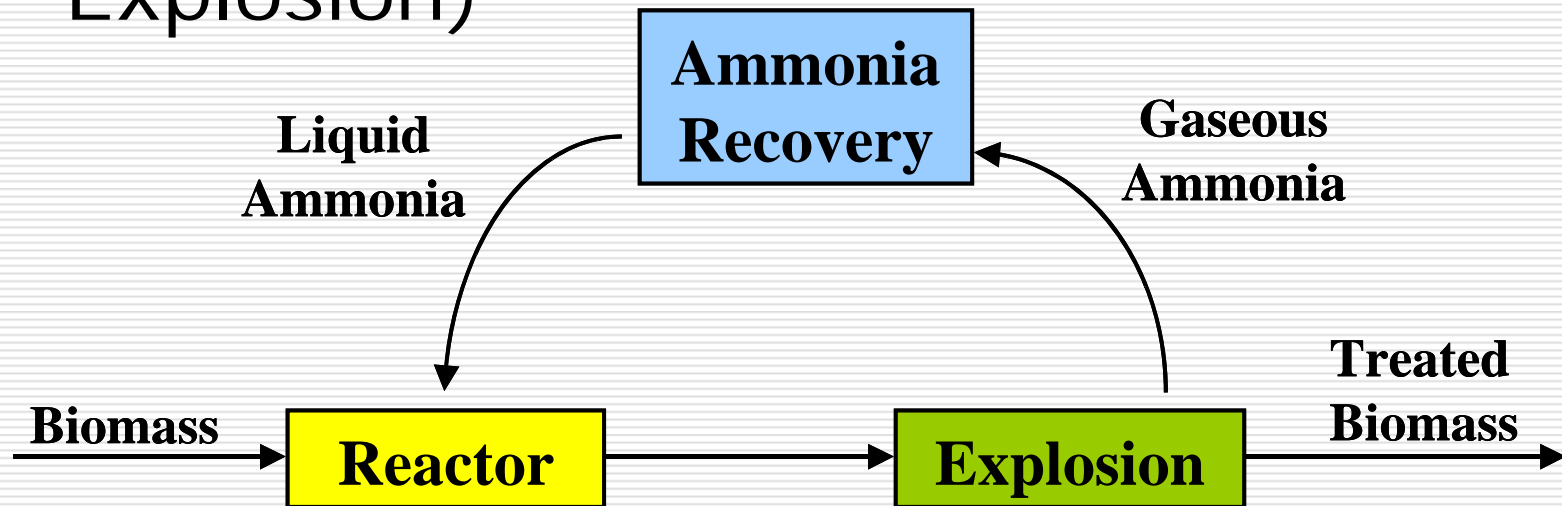
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- Effect of AFEX (Ammonia Fiber Explosion)
- Understanding mechanism of AFEX & effect on enzyme synergy
  - ESCA (Electron Spectroscopy for Chemical Analysis)
  - Prussian Blue Characterization
  - Effect of particle size on AFEX
  - Washed vs. Unwashed Biomass Hydrolysis
- Enzyme Synergy for Lignocellulosics
- Micro-plate Experiments for “Enzyme Cocktail” Optimization



# PART 1

## □ EFFECT OF AFEX (Ammonia Fiber Explosion)



### Typical AFEX Conditions

Pressure (20-30 atm),  
Temperature (60-120 °C),

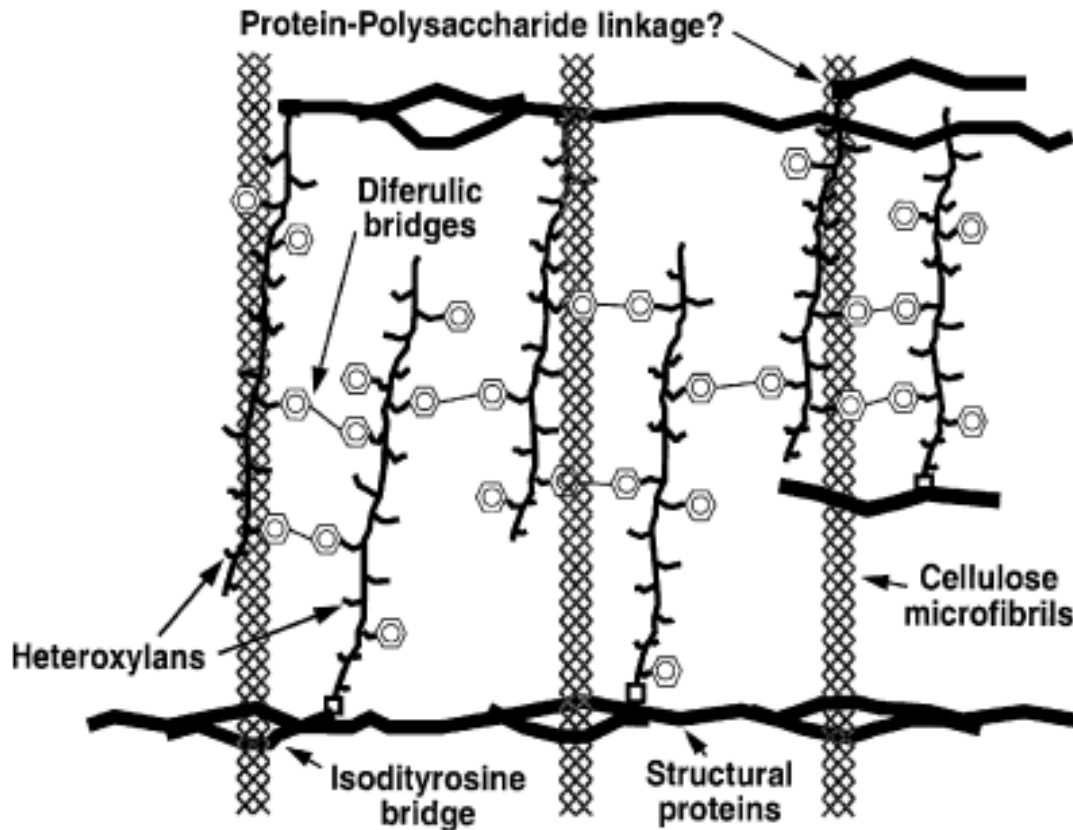
Time (5-15 mins)



# Removal of Alkali Solubles by AFEX

## □ AFEX Effect on

- Acetyl Groups
- Hemicellulose
- Cellulose
- LCC Bonds
- Lignin
- Proteins
- Ca as binding agent for Uronic acids/ Pectins etc.

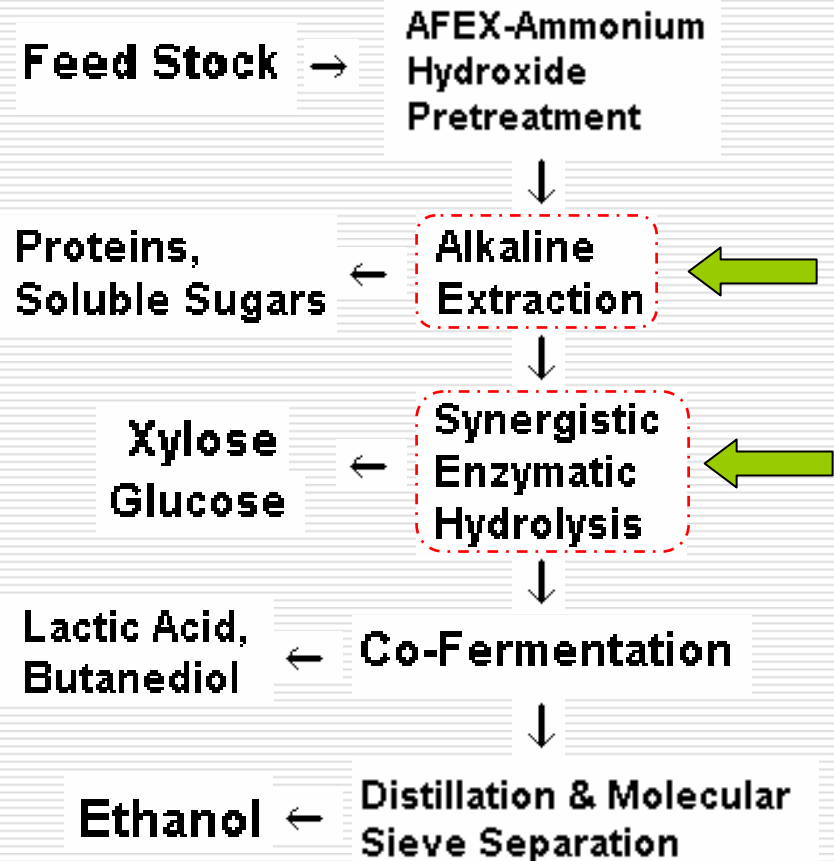


Corn Fiber Cell Wall Model

# AFEX Integrated Biorefinery

## Key Points:

- Relevance of washing/alkali extraction on the synergistic hydrolysis
- Water Washing Protocol  
10% Biomass in water  
Stir for 15 mins  
Drain/Squeeze till dry



Integrated Biorefinery  
Unit Operations



# Types of Biomass Studied

1. Biomass Samples (Corn Stover, Poplar, DDG)
2. Corn Stover Mesh Cut Notation & Composition \*

Sample Notation	Size (microns)	Glucan	Xylan	Galactan	Arabinan	Mannan	Lignin	Protein	Water Solubles	Alcohol Solubles
uncut CS	> 850	32.58	23.44	1.37	2.74	0.26	12.27	3.13	3.89	3.53
# 20	850	33.45	27.52	1.02	2.35	0.69	12.46	1.75	0.14	2.28
# 35	500	34.35	24.73	1.31	2.68	0.38	12.37	2.01	2.68	3.15
# 40	425	34.42	23.54	1.27	2.52	0.30	12.63	2.25	4.31	3.24
# 70	212	33.99	22.58	1.40	2.62	0.21	12.59	2.71	5.98	3.62
# 80	180	33.99	21.85	1.51	2.66	0.07	12.94	2.89	7.91	3.85
# 100	150	33.33	21.92	1.66	2.89	0.05	12.82	3.27	8.51	3.90
# 120	<150	32.29	20.06	1.75	2.93	0.00	12.01	4.27	11.28	4.37

# PART 2

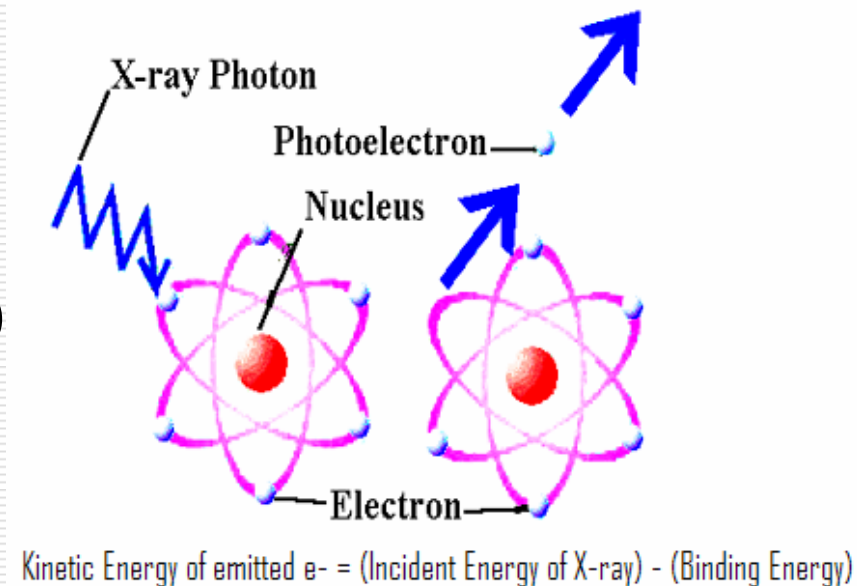
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- UNDERSTANDING EFFECT OF AFEX VIA VARIOUS TECHNIQUES
  - ESCA
  - PRUSSIAN BLUE
  - PARTICLE SIZE
  - WATER WASHING



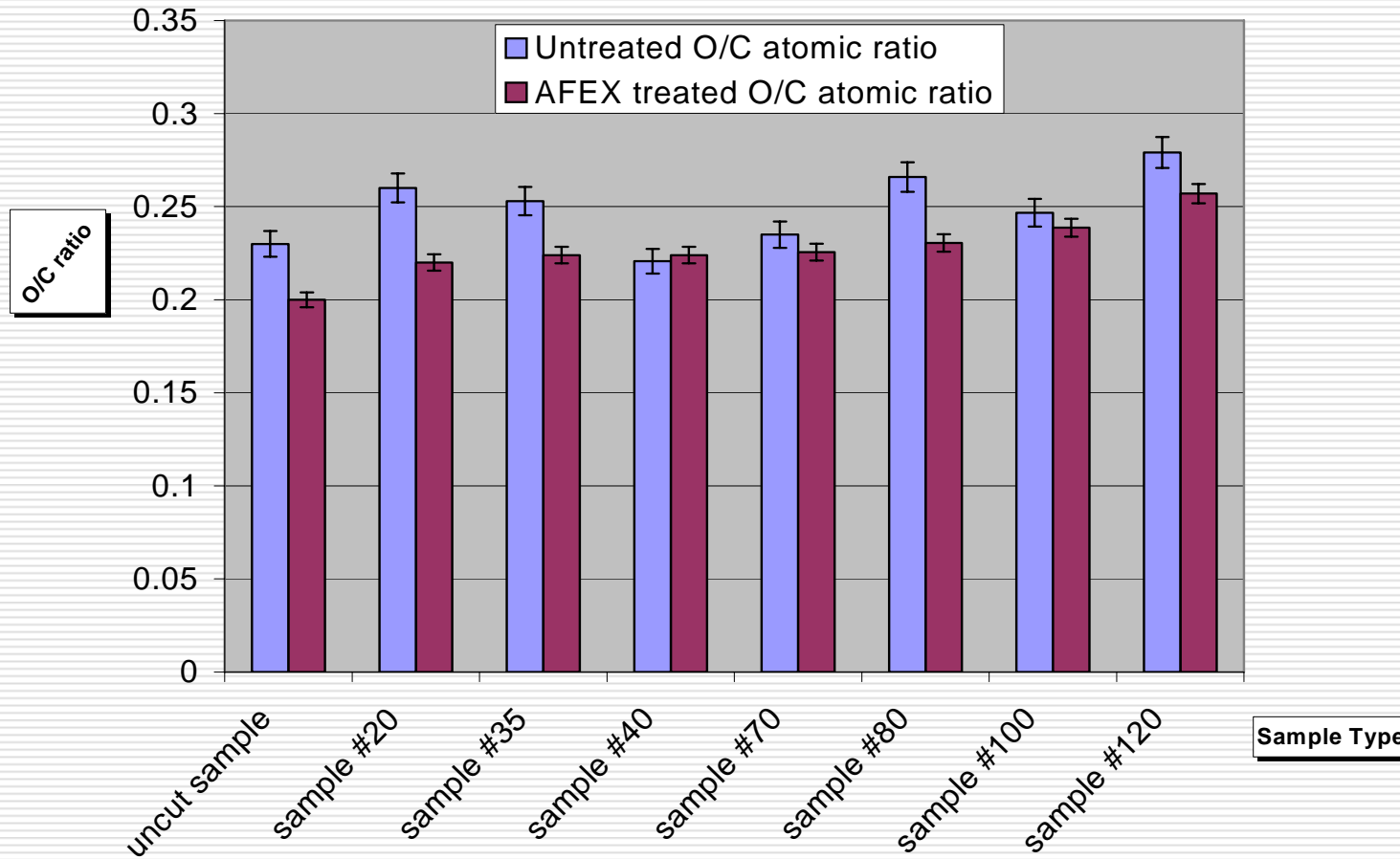
# ESCA as a tool to study effect of AFEX

- ESCA or XPS (X-ray Photoelectron Spectroscopy) Principles
  - Surface Analysis (depth upto 1-20 nm)
  - Types of Chemical Bonding (C,O,N,Ca)
  - O/C ratio
    - Eg. (Gustafsson, 2003)
    - Cellulose O/C = 0.8
    - Lignin O/C = 0.33



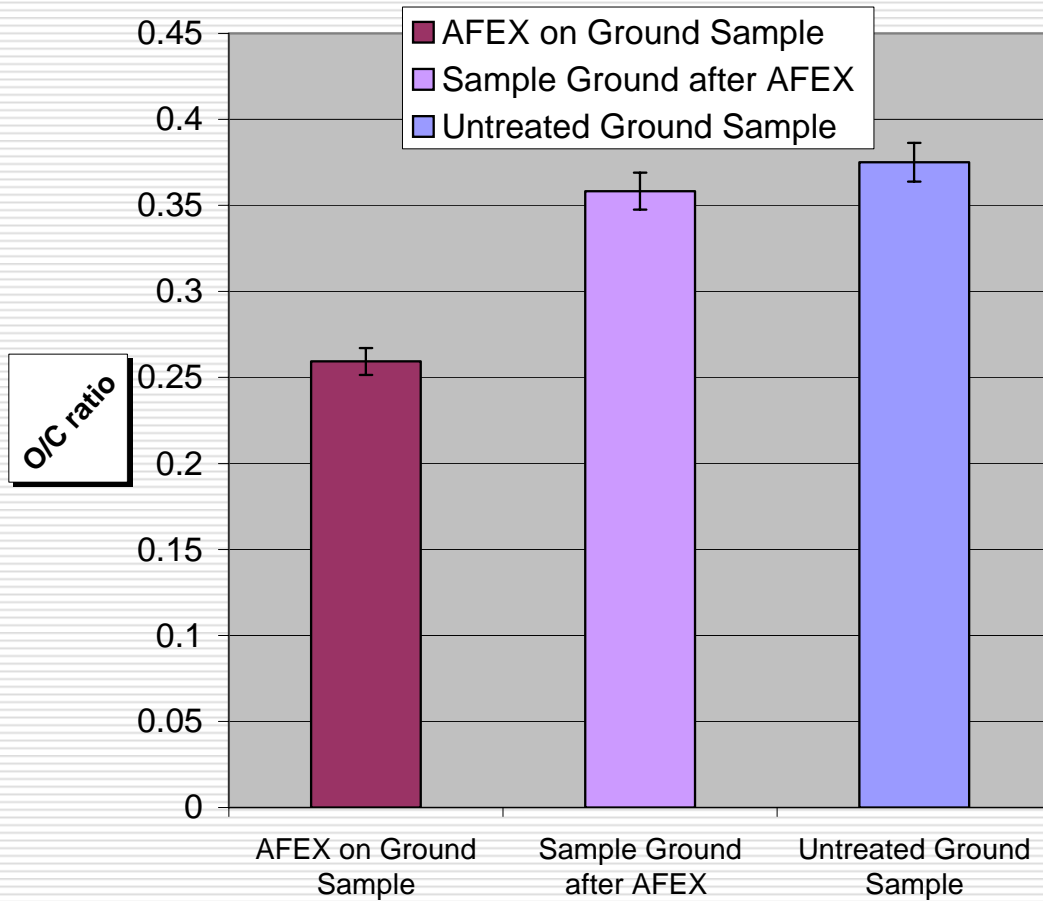
# ESCA O/C ratio for Corn Stover Mesh Cuts

Atomic Ratio of Oxygen (O) to Carbon (C)



# Effect of Size on AFEX

O/C ratio for AFEX treatment before and after grinding corn stover to 80 microns



**Reducing particle size aids in extraction of lignin cleavage fragments & other alkali extractives to the surface**



# ESCA Results Summary

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- ❑ Nitrogen & Calcium signal increased substantially after the AFEX for Corn Stover
- ❑ 5-15 % drop in O/C ratio after AFEX for Corn Stover
- ❑ 30 % drop in O/C ratio after AFEX on Ground Corn Stover



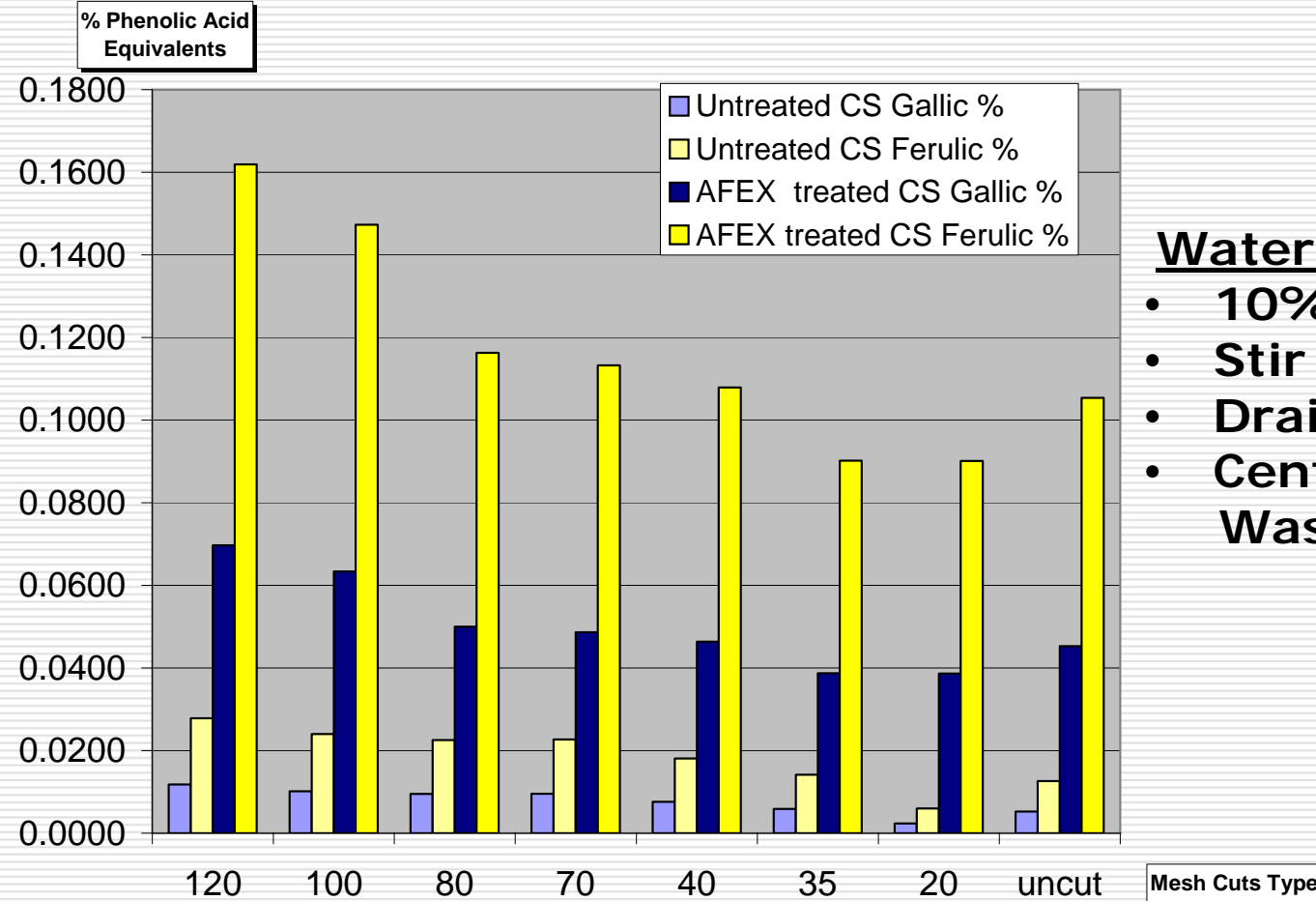
# Modified Prussian Blue Test

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- ❑ Oxidation of phenolic groups...coupled to the reduction of reagent
- ❑ Quantitative Colorimetric Method
- ❑ Standard Phenolic Compounds
  - Gallic Acid & Ferulic Acid
- ❑ Stability of Color
- ❑ High Reproducibility

# Phenolic Content of Wash Streams

**Untreated vs. AFEX Wash Stream Phenolic Content**



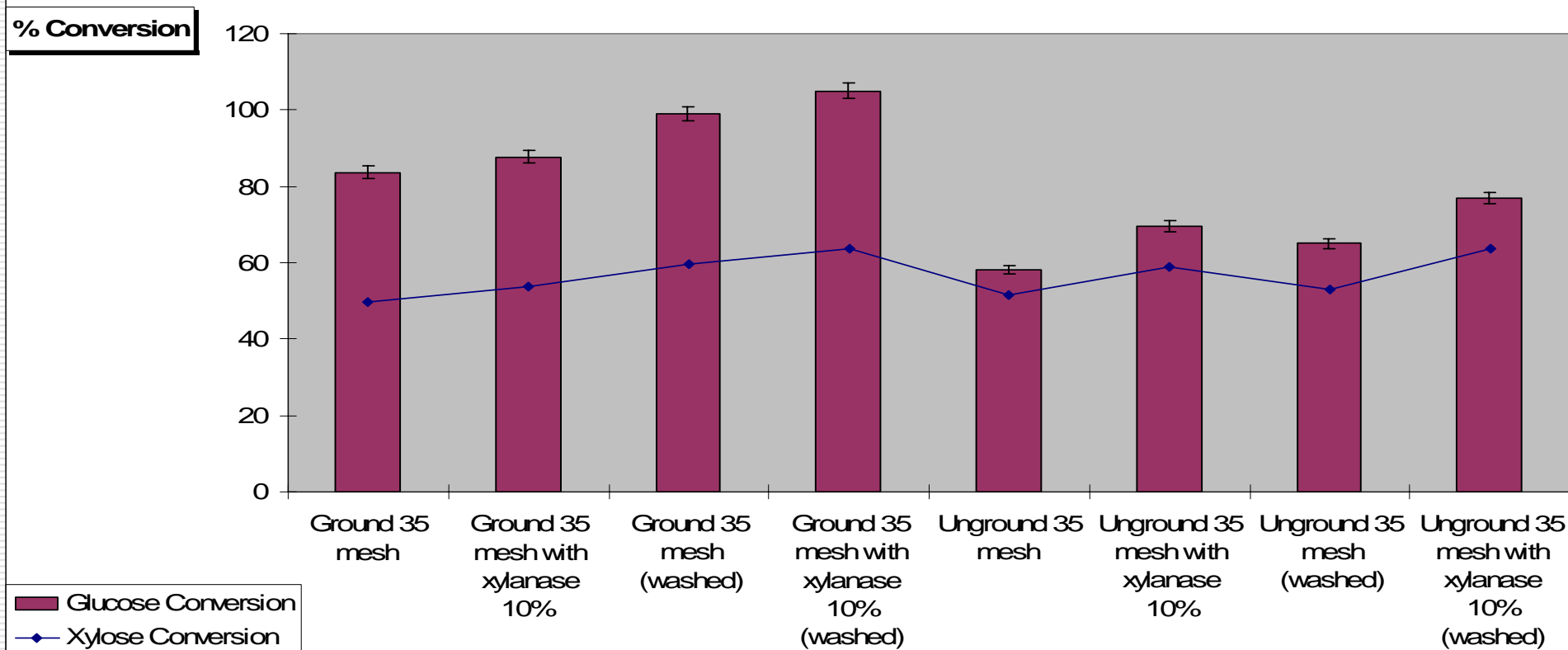
## Water Washing Protocol

- 10% Biomass in water
- Stir for 15 mins
- Drain/Squeeze till dry
- Centrifuge & Filter Wash Liquid



# Hydrolysis Data for Washed vs. Unwashed 35 Mesh Cut

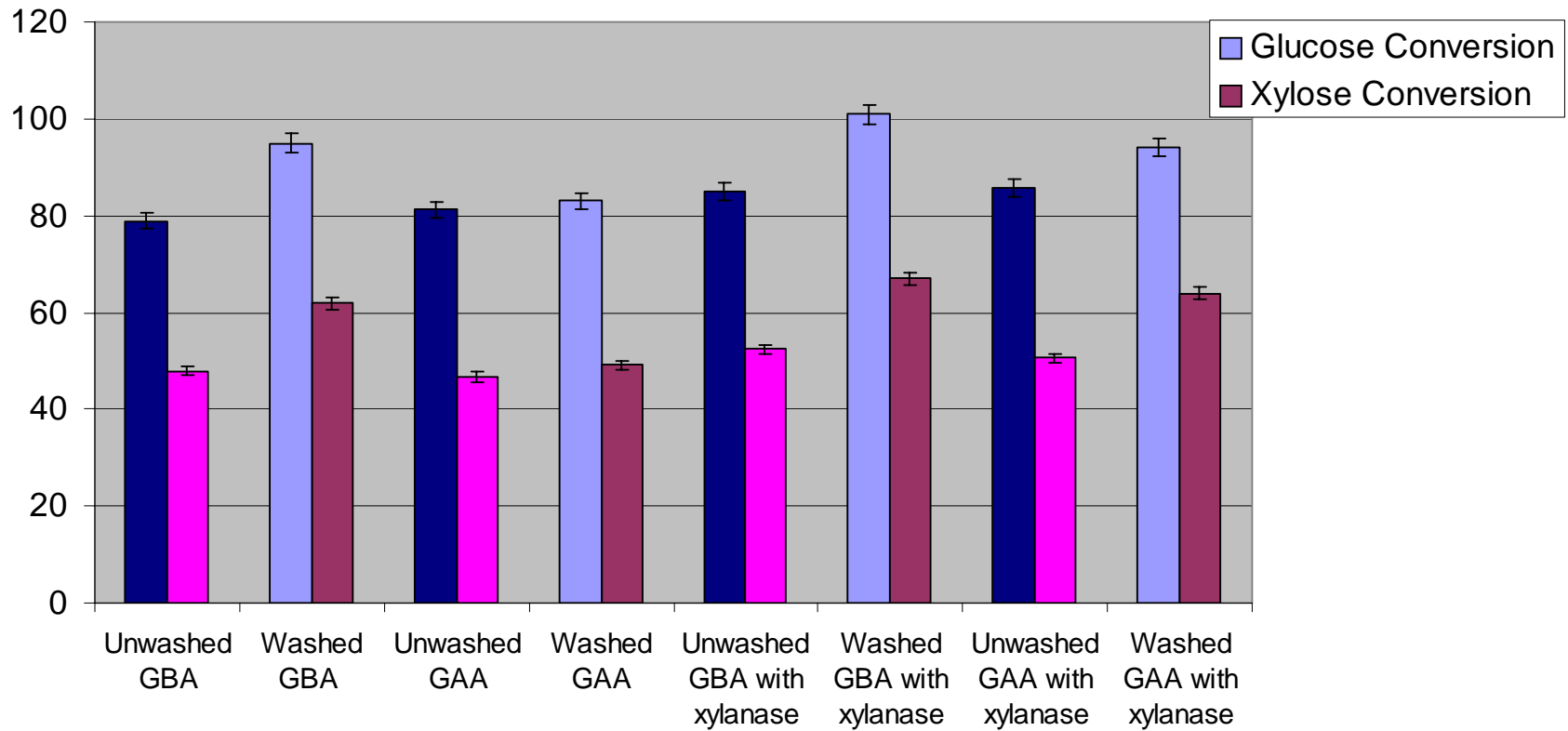
**72 hour Hydrolysis data for AFEX treated CS (Washed/Unwashed)**



NOTE. Unground 35 Mesh Cut (500 microns)  
Ground 35 Mesh Cut (<100 microns)

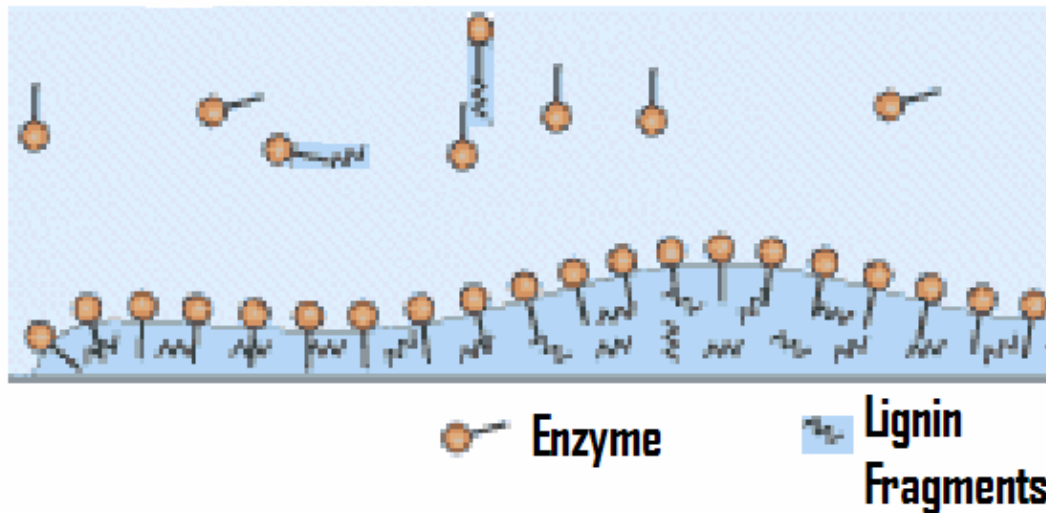
Effect of particle size on AFEX alkali extraction (72 hour hydrolysis data)

Conversion



# Possible Effect of Lignin Fragments extracted during AFEX

## Lignocellulose Surface Modification after AFEX



- This might explain why washing aids in the hydrolysis of biomass, as some of these phenolic fragments are removed.
- What would be the next step is to couple alkali extracted biomass (for protein removal) with the AFEX process.

# PART 3

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## □ SYNERGISTIC HYDROLYSIS FOR LIGNOCELLULOSIC BIOMASS



# Synergistic Hydrolysis

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- **Need to study synergy for lignocellulosic hydrolysis**
  - Role of Lignin and influence of alkali extraction
  - Optimizing multi-enzyme systems could aid the enzyme companies in preparing tailor made enzymes
    - lack of hemicellulase activity,
    - specific cocktails for different pretreatments,
    - cocktail optimization for various biomass compositions,
    - Multiple enzyme cocktails for SSF/SSCF systems
  - Reducing net amount of enzymes (synergy, additives)
  - Reducing hydrolysis time (alkali/water extraction, size reduction)



# PART 4

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- DEVELOPMENT OF MICROPLATE TECHNIQUE FOR “ENZYMES COCKTAIL SCREENING”



# Advantages of Microplate Experiments

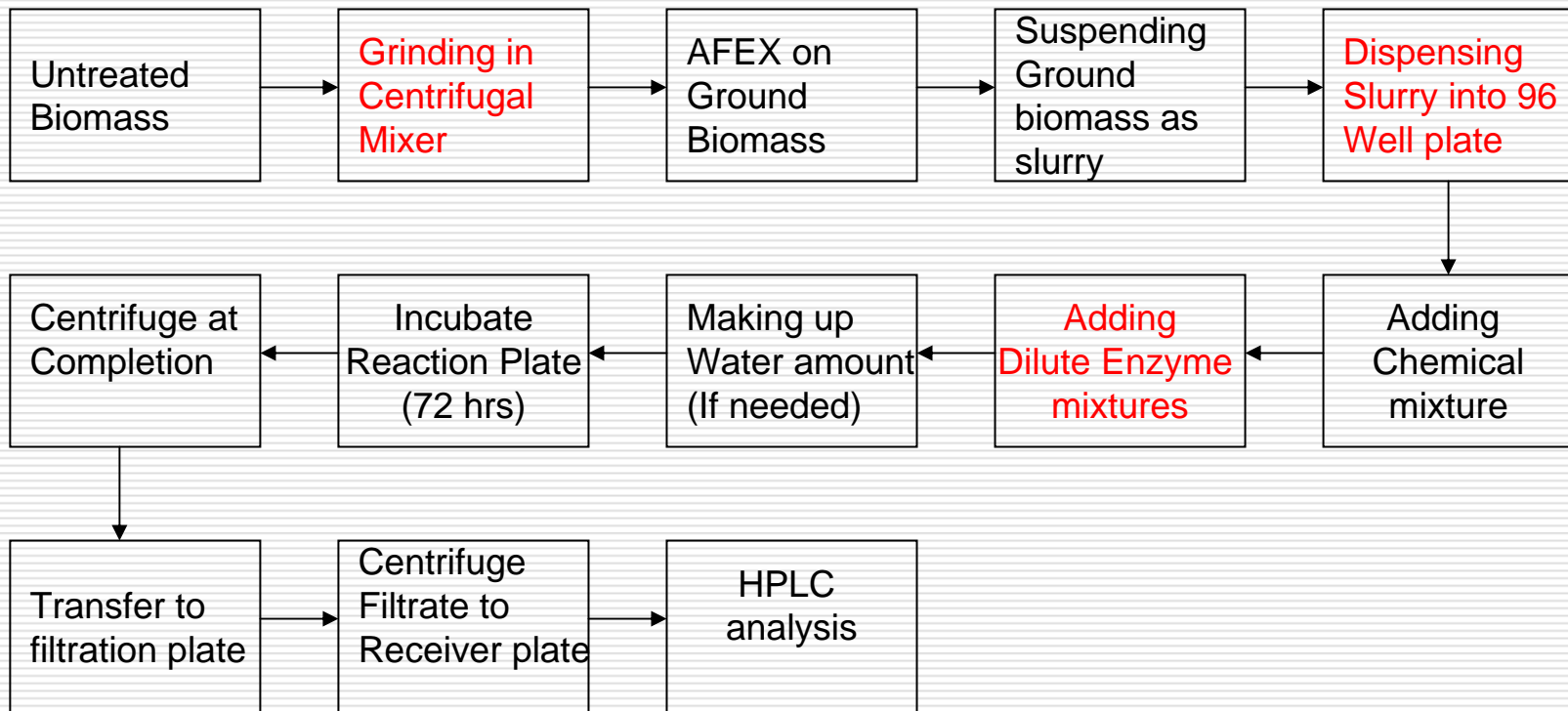
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**Ultimate aim** – analyze hydrolysis profile for multi enzyme systems rapidly over a *larger gamut of input variables*, much *faster* than the conventional protocol in a *reproducible manner*.

- AFEX conditions (better screening of AFEX conditions)
- Enzyme cocktails & concentrations
- Substrate concentration
- Additives
- Protein Extraction (Integrated Step)



# Process Flowchart for Microplate Enzymatic Digestion



# Preliminary Microplate Results

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- Measure enzymatic hydrolysis of cellulose and hemicellulose as a function of cellulase and xylanase and beta glucosidase loadings for Corn Stover
- Determine key trends and synergistic interactions between enzymes



# Continuing Work

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- ❑ AFEX achieved slightly higher yields from enzymes even though no lignin was removed in comparison to other pretreatments and hence need to study the lignin fragmentation due to AFEX
- ❑ Analysis of wash stream sugar/xylooligosaccharides/lignin
- ❑ Fermentability of washed/unwashed biomass streams
- ❑ Inhibition of enzymes for washed/unwashed biomass
- ❑ Microplate Experiments for various enzyme combinations



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- ❑ Genencor
- ❑ MSU BCRL Group



# Questions

## Biomass Conversion Research Group at MSU



# References

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- Saulnier L, Thibault JF (1999) Ferulic acid and diferulic acids as components of sugar-beet pectins and maize bran heteroxylans. *J Sci Food Agric* 79:396–402
- Graham, H. D. (1992) *J Agric Food Chem* 40:801-805

